Clinical Track

Protein-Truncating Variants at the Cholesteryl Ester Transfer Protein Gene and Risk for Coronary Heart Disease

Akihiro Nomura, Hong-Hee Won, Amit V. Khera, Fumihiko Takeuchi, Kaoru Ito, Shane McCarthy, Connor A. Emdin, Derek Klarin, Pradeep Natarajan, Seyedeh M. Zekavat, Namrata Gupta, Gina M. Peloso, Ingrid B. Borecki, Tanya M. Teslovich, Rosanna Asselta, Stefano Duga, Piera A. Merlini, Adolfo Correa, Thorsten Kessler, James G. Wilson, Matthew J. Bown, Alistair S. Hall, Peter S. Braund, David J. Carey, Michael F. Murray, H. Lester Kirchner, Joseph B. Leader, Daniel R. Lavage, J. Neil Manus, Dustin N. Hartze, Nilesh J. Samani, Heribert Schunkert, Jaume Marrugat, Roberto Elosua, Ruth McPherson, Martin Farrall, Hugh Watkins, DiscovEHR Study Group, Jyh-Ming J. Juang, Chao A. Hsiung, Shih-Yi Lin, Jun-Sing Wang, TAICHI Consortium, Hayato Tada, Masa-aki Kawashiri, Akihiro Inazu, Masakazu Yamagishi, Tomohiro Katsuya, Eitaro Nakashima, Masahiro Nakatochi, Ken Yamamoto, Mitsuhiro Yokota, Yukihide Momozawa, Jerome I. Rotter, Eric S. Lander, Daniel J. Rader, John Danesh, Diego Ardissino, Stacey Gabriel, Cristen J. Willer, Goncalo R. Abecasis, Danish Saleheen, Michiaki Kubo, Norihiro Kato, Yii-Der Ida Chen, Frederick E. Dewey, Sekar Kathiresan

<u>Rationale:</u> Therapies that inhibit CETP (cholesteryl ester transfer protein) have failed to demonstrate a reduction in risk for coronary heart disease (CHD). Human DNA sequence variants that truncate the *CETP* gene may provide insight into the efficacy of CETP inhibition.

<u>Objective:</u> To test whether protein-truncating variants (PTVs) at the *CETP* gene were associated with plasma lipid levels and CHD.

Methods and Results: We sequenced the exons of the CETP gene in 58469 participants from 12 case-control studies (18817 CHD cases, 39652 CHD-free controls). We defined PTV as those that lead to a premature stop, disrupt canonical splice sites, or lead to insertions/deletions that shift frame. We also genotyped 1 Japanese-specific PTV in 27561 participants from 3 case-control studies (14286 CHD cases, 13275 CHD-free controls). We tested association of CETP PTV carrier status with both plasma lipids and CHD. Among 58469 participants with CETP gene-sequencing data available, average age was 51.5 years and 43% were women; 1 in 975 participants carried a PTV at the CETP gene. Compared with noncarriers, carriers of PTV at CETP had higher high-density lipoprotein cholesterol (effect size, 22.6 mg/dL; 95% confidence interval, 18–27; P<1.0×10⁻⁴), lower low-density lipoprotein cholesterol (-12.2 mg/dL; 95% confidence interval, -23 to -0.98; P=0.033), and lower triglycerides (-6.3%; 95% confidence interval, -12 to -0.22; P=0.043). CETP PTV carrier status was associated with reduced risk for CHD (summary odds ratio, 0.70; 95% confidence interval, 0.54–0.90; P=5.1×10⁻³).

<u>Conclusions:</u> Compared with noncarriers, carriers of PTV at *CETP* displayed higher high-density lipoprotein cholesterol, lower low-density lipoprotein cholesterol, lower triglycerides, and lower risk for CHD. (*Circ Res.* 2017;121:81-88. DOI: 10.1161/CIRCRESAHA.117.311145.)

Key Words: case-control studies ■ cholesteryl ester transfer protein ■ coronary disease ■ lipids

In 3 randomized controlled trials (RCTs), therapies that inhibit CETP (cholesteryl ester transfer protein) have failed to demonstrate a reduction in risk for coronary heart disease (CHD).¹⁻³ Possible reasons for this failure include ontarget lack of efficacy, off-target adverse effects of the small

In This Issue, see p 2 Meet the First Author, see p 3

molecule, and RCT design factors, such as insufficient statistical power, concurrent statin therapy, or selection of study

Original received April 11, 2017; revision received April 28, 2017; accepted May 12, 2017. In April 2017, the average time from submission to first decision for all original research papers submitted to *Circulation Research* was 11.94 days.

For the author affiliations, see the Appendix.

The online-only Data Supplement is available with this article at http://circres.ahajournals.org/lookup/suppl/doi:10.1161/CIRCRESAHA. 117.311145/-/DC1.

Correspondence to Sekar Kathiresan, MD, Center for Genomic Medicine, Massachusetts General Hospital, 185 Cambridge St, CPZN 5.830, Boston, MA 02114. E-mail skathiresan1@mgh.harvard.edu

© 2017 American Heart Association, Inc.

Circulation Research is available at http://circres.ahajournals.org

Novelty and Significance

What Is Known?

- Human DNA sequence variants that truncate a therapeutic target protein may provide insight into the efficacy of pharmacological inhibition.
- It has been uncertain whether carriers of protein-truncating variants (PTVs) at the cholesteryl ester transfer protein (*CETP*) gene have altered plasma lipid levels and lower risk for coronary heart disease (CHD).

What New Information Does This Article Contribute?

- Carriers of a PTV at CETP had higher high-density lipoprotein cholesterol, lower low-density lipoprotein cholesterol, and lower triglycerides.
- CETP PTV carrier status was also associated with 30% reduced risk for CHD.
- Lifelong reduction in CETP function is associated with altered plasma lipids and a lower risk for CHD.

Therapies that inhibit CETP have failed to demonstrate a reduction in risk for CHD. Human DNA sequence variants that truncate

a therapeutic target gene may provide insight into the efficacy of pharmacological inhibition. We tested whether humans carrying PTVs at the CETP gene were associated with lipid levels and were at reduced risk for CHD. We sequenced the exons of the CETP gene in 58 469 participants from 12 case-control and genotyped 1 Japanese-specific PTV in 27561 participants from 3 case-control studies. PTVs at the CETP gene were defined as mutations that lead to a premature stop, disrupt canonical splice sites, or lead to insertions/deletions that shift frame. In an analysis including >80 000 participants, carriers of a PTV at CETP had higher high-density lipoprotein cholesterol (+22.6 mg/dL), lower low-density lipoprotein cholesterol (-12.2 mg/dL), and lower triglycerides (-6.3%). CETP PTV carrier status was also associated with 30% reduced risk for CHD (summary odds ratio, 0.70). In conclusion, compared with noncarriers, carriers of PTV at the CETP gene displayed higher high-density lipoprotein cholesterol, lower low-density lipoprotei cholesterol, lower triglycerides, and lower risk for CHD.

Nonstandard Abbreviations and Acronyms

BBJ BioBank Japan

CAGE-CAD Cardio-metabolic Genome Epidemiology Network and

Coronary Artery Disease

CETP cholesteryl ester transfer protein

CHD coronary heart disease

CI confidence interval
HDL-C high-density lipoprotein cholesterol

LDL-C low-density lipoprotein cholesterol
MIGen Myocardial Infarction Genetics

PCSK9 proprotein convertase subtilisin/kexin type 9

PTV protein-truncating variant
RCT randomized controlled trial

participants. 4-6 An RCT of a fourth CETP inhibitor—anacetrapib—is ongoing. 7

Studies of humans with naturally occurring genetic variation in genes encoding drug targets can provide insight into the potential efficacy and safety of therapeutic modulation targeting the gene product.^{8–10} Genetic studies of common, regulatory variants at the *CETP* gene region initially showed mixed results^{11–15} but more recently have converged on a consensus finding: alleles with lower CETP expression are associated with reduced CHD risk.¹⁶

Beyond common DNA sequence variants, rare mutations that truncate a therapeutic target gene may be of particular value because they most closely mirror pharmacological inhibition. 8,9,17 Protein-truncating variants (PTVs; ie, nonsense, canonical splice site, and frameshift mutations) at 2 therapeutic targets—NPC1L1 (NPC1-like intracellular cholesterol transporter 1)9 and proprotein convertase subtilisin/kexin type 9 (PCSK9)8—are associated with lower low-density lipoprotein cholesterol (LDL-C) and reduced CHD risk. A therapeutic trial testing NPC1L1 inhibition was consistent with the human genetic findings, 18 and a

trial testing PCSK9 inhibition was consistent as well.¹⁹ Here, we tested whether rare PTVs at the *CETP* gene were associated with plasma lipids and reduced odds of CHD.

Methods

Study Participants

First, we sequenced a total of 58469 participants from the MIGen consortium (Myocardial Infarction Genetics) of African, European, and South Asian ancestries (n=25273), the DiscovEHR (DiscovEHR project of the Regeneron Genetics Center and the Geisinger Health System) of European ancestry (n=24138),²⁰ and Taiwanese-Chinese (TAICHI) consortium of East Asian ancestry (n=9058)²¹ (Table 1). The MIGen consortium consists of the Italian ATVB study (Atherosclerosis Thrombosis and Vascular Biology),22 the DHM study (Deutsches Herzzentrum München Myocardial Infarction),9 the ESP-EOMI study (Exome Sequencing Project Early-Onset Myocardial Infarction)^{23,24} of European and African ancestries, the JHS (Jackson Heart Study),25 the Leicester (Leicester Acute Myocardial Infarction Peptide Study),²⁶ the Lubeck (Lübeck Myocardial Infarction Study),²⁷ the OHS (Ottawa Heart Study),²⁸ the PROCARDIS (Precocious Coronary Artery Disease Study),²⁹ the PROMIS (Pakistan Risk of Myocardial Infarction Study),³⁰ and the REGICOR (Registre Gironi del COR) study.³¹

We also genotyped a Japanese-specific PTV at the *CETP* gene (rs5742907; IVS14+1G>A; splice-donor variant³²) in a total of 27 561 Japanese participants from BioBank Japan (BBJ)³³ and the CAGE-CAD (Cardio-metabolic Genome Epidemiology Network and Coronary Artery Disease) stage 1 and stage 2 studies (Table 1).³⁴

All participants in the study provided written informed consent for genetic studies. The Institutional Review Boards at the Broad Institute and each participating institution approved the study protocol.

Definition of CETP PTVs

PTVs were defined as premature stop (nonsense), canonical splice sites (splice donor or splice acceptor), including IVS14+1G>A (rs5742907), or insertion/deletion variants that shifted frame (frameshift). The positions of these PTVs were based on the GRGh37 human genome reference and the canonical transcript for *CETP* (transcript ID: ENST00000200676).

Clinical Characteristics, Lipid Measurements, and Definition of CHD

A medical history and laboratory data for cardiovascular risk factors were obtained from all the study participants. Plasma total cholesterol,

Table 1. Clinical Characteristics of Each Study by Protein-Truncating Variant Carrier Status

	NA.	ICon	DingovELID		TAICH		DD I		CACE CAD Ctogo 1		CACE CAD Ctore 2	
	MIGen		DiscovEHR		TAICHI		BBJ		CAGE-CAD Stage 1		CAGE-CAD Stage 2	
	PTV Carrier	Noncarrier	PTV Carrier	Noncarrier	PTV Carrier	Noncarrier	PTV Carrier	Noncarrier	PTV Carrier	Noncarrier	PTV Carrier	Noncarrier
	n=26	n=25 247	n=16	n=24122	n=18	n=9040	n=122	n=15476	n=21	n=2584	n=76	n=9282
Age, y, mean (SD)	53.5 (13)	53.3 (13)	47.2 (12)	46.2 (12)	61.2 (15)	60.9 (15)	65.1 (10)	65.2 (10)	66.4 (8)	65.9 (8)	63.6 (8)	62.5 (7)
Male gender, n (%)	26 (79)	18 387 (73)	9 (56)	5777 (24)	12 (67)	6166 (68)	78 (64)	10 943 (71)	12 (57)	1711 (66)	48 (63)	6041 (65)
BMI, kg/m², median (IQR)	25.7 (23–28)	26.2 (24–29)	33.7 (31–38)	31.2 (26–37)	24.8 (23–30)	24.9 (23–28)	23.2 (21–25)	23.4 (21–26)	23.6 (21–24)	23.3 (21–25)	23.7 (22–26)	23.2 (21–24)
Current smoker, n (%)	7 (21)	7389 (29)	3 (19)	5048 (21)	N/A	N/A	78 (64)	10 282 (66)	11 (52)	1336 (51)	36 (47)	4752 (51)
Medical history												
Hypertension, n (%)	12 (36)	9499 (38)	7 (44)	12 933 (54)	6 (33)	4783 (53)	54 (44)	6408 (41)	6 (28)	1182 (45)	39 (51)	4760 (51)
Type 2 diabetes mellitus, n (%)	8 (24)	5069 (20)	5 (31)	4126 (17)	8 (44)	4343 (48)	78 (64)	8568 (55)	9 (42)	917 (35)	16 (21)	2174 (23)
Lipid-lowering medication,* n (%)	1 (3)	3682 (15)	4 (25)	6129 (25)	3 (19)	1781 (22)	43 (35)	5164 (33)	6 (28)	377 (14)	N/A	N/A
Lipid profile												
LDL cholesterol, mean (SD)	121 (57)	130 (48)	114 (39)	124 (38)	126 (42)	120 (50)	118 (35)	125 (38)	117 (32)	130 (38)	N/A	N/A
HDL cholesterol, mean (SD)	61 (24)	41 (14)	58 (14)	51 (15)	58 (23)	45 (14)	67 (25)	50 (15)	78 (24)	58 (16)	N/A	N/A
Triglycerides, median (IQR)	124 (70–163)	150 (102–222)	162 (105–198)	126 (89–177)	138 (89–186)	121 (83–176)	138 (89–156)	145 (86–175)	74 (57–131)	109 (80–154)	N/A	N/A
Total cholesterol, mean (SD)	211 (61)	206 (54)	207 (50)	205 (42)	210 (45)	190 (46)	239 (72)	231 (61)	218 (37)	214 (41)	N/A	N/A

BBJ indicates BioBank Japan; BMI, body mass index; CAGE-CAD, the Cardio-metabolic Genome Epidemiology Network and Coronary Artery Disease study; DiscovEHR, the DiscovEHR project of the Regeneron Genetics Center and the Geisinger Health System; HDL, high-density lipoprotein; IQR, interquartile range; LDL, low-density lipoprotein; MIGen, Myocardial Infarction Genetics Consortium; N/A, not applicable; PTV, protein-truncating variant; and TAICHI, Taiwanese-Chinese consortium.

triglycerides, and high-density lipoprotein cholesterol (HDL-C) levels were determined enzymatically. LDL-C level was calculated using the Friedewald equation^{35,36} for those with triglycerides <400 mg/dL. If triglycerides ≥400 mg/dL, LDL-C level was directly measured or set to missing. The effect of lipid-lowering therapy at the time of lipid measurement was taken into account by dividing the measured total cholesterol and LDL-C levels by 0.8 and 0.7, respectively.³⁷ HDL-C and triglyceride levels were not adjusted by lipid-altering medication use, and triglyceride levels were natural logarithm transformed for statistical analysis. CHD case and CHD-free control definitions of each study are in Online Table I.

Sequencing and Genotyping to Characterize PTVs

Whole-exome sequencing of the MIGen consortium was performed at the Broad Institute (Cambridge, MA) as previously described.²³ Sequencing reads were aligned to a human reference genome (build 37) using the Burrows–Wheeler Aligner-maximal exact match algorithm. Aligned nonduplicate reads were locally realigned, and base qualities were recalibrated using the Genome Analysis ToolKit software.³⁸ Variants were jointly called using the Genome Analysis ToolKit HaplotypeCaller program. The sensitivity of the Variant Quality Score Recalibration threshold was 99.6% for single nucleotide variants and 95% for insertion/deletion variants. All identified variants were annotated with the use of the Variant Effect Predictor software (version 82).³⁹ The DiscovEHR project and TAICHI consortium participants were exome sequenced as previously described.²⁰

We also genotyped 1 splice-donor variant (IVS14+1G>A [rs5742907]) at the *CETP* gene using the multiplex polymerase chain reaction–based target sequencing in BBJ⁴⁰ or the TaqMan assay in CAGE-CAD stage 1 and stage 2.

Statistical Analysis

We tested the association of *CETP* PTV carrier status with lipid levels using linear regression adjusted by age, sex, study, and the first 5 principal components of ancestry (MIGen), or by age and sex (BBJ and CAGE-CAD stage 1). Only CHD-free controls in each study were included in this assessment to minimize the effect of ascertainment bias. These data were meta-analyzed to calculate overall summary effect sizes with an inverse-variance weighted fixed-effects model.

We tested the association of *CETP* PTV carrier status with CHD risk using a Cochran–Mantel–Haenszel method without continuous correction. This method combines score statistics instead of Wald statistics and is useful for rare exposures when some observed odds ratios are zero. We removed ESP-EOMI and JHS from this analysis because no participant in these 2 studies carried a PTV at the *CETP* gene.

In an exploratory analysis, we evaluated whether the effect of *CETP* PTVs on LDL-C could explain the reduction in CHD risk. We used an inverse-variance weighted model to draw a regression line with a 95% confidence interval (CI). Across 4 genes (*APOB*, *NPC1L1*, *PCSK9*, and *CETP*), we plotted the effect of DNA sequence variants in these genes on both LDL-C and CHD risk. The results for *APOB*, *NPC1L1*, and *PCSK9* are derived from samples of the MIGen

^{*}At the time of lipid measurement.

consortium to draw a dose-response reference line. The results for *CETP* are summary estimate from all studies.

Statistical analyses were performed using R software version 3.2.3 (The R Project for Statistical Computing, Vienna, Austria).

Results

Prevalence of CETP PTVs

Sequencing of the 16 exons at the *CETP* gene was performed in 58469 participants (18817 CHD cases and 39652 CHD-free controls) from 3 projects: the MIGen consortium, the DiscovEHR project, and TAICHI consortium. Baseline characteristics of each study are shown in Table 1. A total of 23 PTVs were identified (10 premature stop, 9 frameshifts, 3 splice-donor variants, and 1 splice-accepter variant). A total of 60 individuals carried one of the *CETP* PTVs, including 18 CHD cases (0.096%; 95% CI, 0.051%–0.14%) and 42 CHD-free controls (0.11%; 95% CI, 0.074%–0.14%). Baseline characteristics by variant carrier status are shown in Online Table II. We genotyped a Japanese-specific splice-donor variant (IVS14+1G>A [rs5742907]) in 3 studies from Japan and found the carrier frequency to be BBJ, 0.78%; CAGE-CAD stage 1, 0.81%; and CAGE-CAD stage 2, 0.92%.

Association of CETP PTVs With Plasma Lipids

We assessed whether *CETP* PTV carrier status was associated with lipid levels (Table 2; Online Figure). We obtained plasma lipid profiles in 11 205 control participants from the MIGen consortium and 6955 control participants from BBJ and CAGE-CAD stage 1. *CETP* PTV carrier status was associated with increased HDL-C (effect size, 22.6 mg/dL; 95% CI, 18–27; *P*<1×10⁻⁴), decreased LDL-C (–12.2 mg/dL; 95% CI, –23 to –0.98; *P*=0.033), and decreased triglycerides (–6.3%; 95% CI, –12 to –0.22; *P*=0.043).

Association of CETP PTVs With CHD

We evaluated the association of *CETP* PTV carrier status with CHD. Baseline characteristics and lists of *CETP* PTVs by case–control status in each study are shown in Online Table III and Online Table IV. In an analysis including a total of 82 722 participants, *CETP* PTV carrier status was significantly associated with lower risk for CHD (summary odds ratio, 0.70; 95% CI, 0.54–0.90; *P*=5.1×10⁻³; Figure 1).

DNA Sequence Variants, LDL-C, and CHD Risk Across 4 Genes

We explored whether the effect size of CETP PTV on CHD risk was consistent with its effect on LDL-C. We drew a

dose-response reference line for CHD risk as a function of LDL-C change conferred by DNA sequence variants in 3 genes other than *CETP*. DNA sequence variants in *APOB*, *NPC1L1*, and *PCSK9* associated with lower LDL-C also correlated with lower CHD risk. The effect of *CETP* PTV on CHD risk (30% reduction in risk) was consistent with the estimate based on the change in LDL-C (-12.2 mg/dL; Figure 2).

Discussion

Across >80 000 participants, we evaluated whether *CETP* PTVs were associated with lipid levels and risk for CHD. About 1 in 975 participants carried a PTV at the *CETP* gene in sequencing studies, and compared with noncarriers, *CETP* PTV carriers exhibited significantly higher plasma HDL-C levels and lower LDL-C and triglyceride levels. The presence of a *CETP* PTV was also associated with decreased risk for CHD.

This evidence from rare human mutations that disrupt the *CETP* gene is consistent with earlier data on common, regulatory variants at the *CETP* locus. Common variants in *CETP* have been associated with increased HDL-C, decreased LDL-C, decreased triglyceride levels,⁴¹ and reduced risk for CHD.^{13,42–44} And recently, the statistical evidence for association of common *CETP* variants with CHD has exceeded a stringent genome-wide threshold.¹⁶ Exploratory analyses suggest that the effect of *CETP* PTV on lower CHD risk is consistent with lower LDL-C change conferred by these variants.

If human genetics shows loss of *CETP* function mutations to be associated with reduced CHD risk, why have 3 small molecule inhibitors of CETP function all failed to show lower CHD outcomes in randomized clinical trials? Several possibilities emerge. First, this could be because of off-target adverse effects of small molecule inhibitors. Torcetrapib, dalcetrapib, and evacetrapib treatment all led to higher blood pressure in RCTs¹⁻³; torcetrapib also led to hyperaldosteronism.¹

Second, RCT design factors, such as limited statistical power, could play a role.^{4,5} Human genetic evidence is supportive for apolipoprotein B–containing lipoproteins (low-density lipoprotein, triglyceride-rich lipoproteins, lipoprotein[a]) as causal factors for CHD, whereas this is not the case for HDL-C.⁶ As such, any benefit from CETP inhibition may be solely because of the lowering of apolipoprotein B–containing lipoproteins. On background statin therapy, the LDL cholesterol and apolipoprotein B–lowering effect are smaller, as shown in the ACCELERATE trial (Assessment of Clinical Effects of Cholesteryl Ester Transfer Protein Inhibition With Evacetrapib in Patients at a High Risk for Vascular Outcomes).³

 Table 2.
 Associations of CETP Protein-Truncating Variant Carrier Status With HDL Cholesterol, LDL Cholesterol, Triglycerides, and

 Total Cholesterol
 Associations of CETP Protein-Truncating Variant Carrier Status With HDL Cholesterol, LDL Cholesterol, Triglycerides, and

	MIGen			В	BJ and CAGE-CAL)	Overall			
	Effect Size	95% CI	P Value	Effect Size	95% CI	P Value	Effect Size	95% CI	<i>P</i> Value	
HDL cholesterol, mg/dL	19.2	12 to 27	2.5×10 ⁻⁷	24.5	19 to 30	<0.0001	22.6	18 to 27	<0.0001	
LDL cholesterol, mg/dL	-20.2	-45 to 4.8	0.11	-10.2	-23 to 2.4	0.11	-12.2	-23 to -0.98	0.033	
Triglycerides, %	2.8%	-27 to 44	0.16	-6.6%	-12 to -0.43	0.036	-6.3%	-12 to -0.22	0.043	
Total cholesterol, mg/dL	5.2	-23 to 33	0.72	8.6	-8.7 to 26	0.97	7.6	-7.1 to 22	0.31	

BBJ indicates BioBank Japan; CAGE-CAD, the Cardio-metabolic Genome Epidemiology Network and Coronary Artery Disease study; CETP, cholesteryl ester transfer protein; CI, confidence interval; HDL, high-density lipoprotein; LDL, low-density lipoprotein; and MIGen, Myocardial Infarction Genetics consortium.

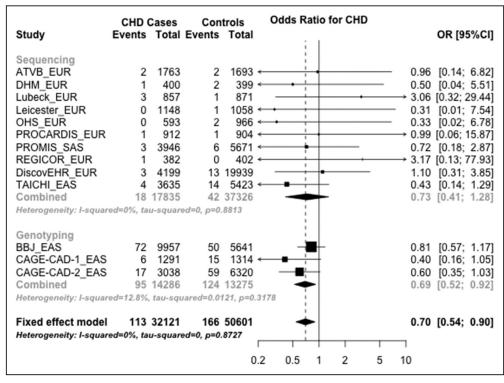


Figure 1. Association of CETP (cholesteryl ester transfer protein) protein-truncating variant carrier status with risk for coronary heart disease (CHD). CETP protein-truncating variant carrier status was associated with reduced risk for CHD. Each study column indicates [Study name] _[Ancestry]. ATVB indicates Atherosclerosis Thrombosis and Vascular Biology; BBJ, BioBank Japan; CAGE-CAD, Cardio-metabolic Genome Epidemiology Network and Coronary Artery Disease; CI, confidence interval; DHM, Deutsches Herzzentrum München Myocardial Infarction Study; DiscovEHR, DiscovEHR project of the Regeneron Genetics Center and the Geisinger Health System; EAS, East Asian ancestry; EUR, European ancestry; OHS, Ottawa Heart Study; OR, odds ratio; PROMIS, Pakistan Risk of Myocardial Infarction Study; REGICOR, Registre Gironi del COR; and SAS, South Asian ancestry.

As such, it is unclear whether RCTs were adequately powered to detect this benefit.

Third, statin therapy may modify the relationship of CETP activity and coronary disease. CETP promotes the transfer of cholesteryl

esters from HDL to atherogenic apolipoprotein B-containing lipoproteins, including LDL.⁴ If not cleared from the circulation, accumulation of such particles in the bloodstream promotes atherosclerotic progression. However, statins lead to substantial

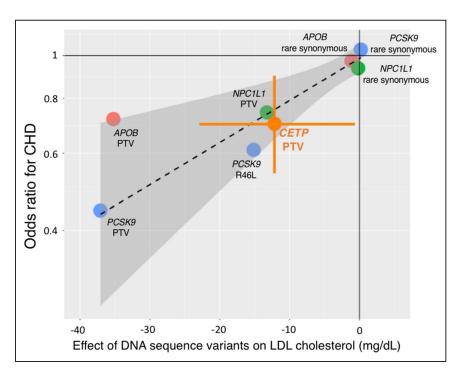


Figure 2. Effects of DNA sequence variants in 4 genes on low-density lipoprotein cholesterol (LDL-C) and coronary heart disease (CHD) risk. Dashed line denotes a dose-response reference line, with the 95% confidence interval (CI) indicated by shadow. Error bar indicates cholesteryl ester transfer protein (CETP) protein-truncating variant (PTV) 95% CIs of an effect size on LDL-C and odds ratio for CHD.

upregulation of hepatic LDL receptor density.⁴⁵ In this context, apolipoprotein B-containing lipoproteins may be rapidly cleared from the circulation and excreted into the feces. CETP may, therefore, play a role in promoting reverse cholesterol transport, the process by which cholesterol is extracted from peripheral tissues (eg, atherosclerotic plaque) and excreted from the body. Overexpression of CETP leads to enhanced reverse cholesterol transport via an LDL receptor-dependent pathway in mouse models.⁴⁶ Furthermore, individuals with increased on-statin CETP mass were protected from recurrent coronary events, particularly when the achieved LDL cholesterol was <80 mg/dL.47 Under this framework, pharmacological CETP inhibition might prove less effective or potentially harmful among those in whom statin therapy leads to efficient clearance of apolipoprotein B-containing lipoproteins. However, the impact of CETP inhibition on reverse cholesterol transport has been questioned because the mouse studies might have been confounded by cholesterol pool size changes.⁴⁸ Also, torcetrapib did not elevate fecal cholesterols or bile acids in both on- and off-statin individuals.49

Fourth, a major difference in individuals with heterozygous PTV mutations in CETP and individuals taking the CETP inhibitors is the accumulation of a cholesteryl ester-rich HDL in the latter, which may be a toxic lipoprotein particle like cholesteryl ester-rich LDL.

Finally, phenotypic consequences of human PTVs reflect lifelong perturbation of a gene in every human tissue. In contrast, the results of RCTs reflect pharmacological inhibition initiated later in life. As such, there are intrinsic limitations in using human mutations to anticipate efficacy and safety of pharmacological manipulation.

These results should be interpreted in the context of study limitations. Definitions of CHD were different among studies. Cases in the MIGen consortium and the DiscovEHR project were limited to only early-onset CHD, whereas those in East Asian studies were not. Loss of *CETP* function alters the distribution of cholesterol and triglycerides in lipoproteins, and as such, LDL-C levels estimated by the Friedewald equation might overestimate the reduction in participants harboring *CETP* PTVs. We only assessed 4 major lipid levels to evaluate effects of *CETP* PTV carrier status, and other traits such as lipoprotein (a) or function of reverse cholesterol transport were unavailable. Results were somewhat stronger in participants from the Japanese genotyping studies, but the point estimates of the odds ratios for CHD were consistent between populations of Japanese and non-Japanese ancestries (0.69 and 0.73, respectively).

Conclusions

In this meta analyses of data from 15 case—control studies, rare PTVs at the *CETP* gene were associated with higher HDL-C, lower LDL-C, lower triglycerides, and reduced risk for CHD.

Acknowledgments

We thank to all the participants and staffs regarding this project. Also, we express our gratitude to Drs Toshihiro Tanaka, Yasuhiko Sakata, and Shinichiro Suna for their contribution to the study discussion.

Sources of Funding

A. Nomura was funded by the Yoshida Scholarship Foundation. H.-H. Won was funded by the National Research Foundation of Korea (NRF) grant funded by the Korea government (Ministry of Science,

ICT and Future Planning; no. 2016R1C1B2007920). A.V. Khera is supported by a KL2/Catalyst Medical Research Investigator Training award from Harvard Catalyst funded by the National Institutes of Health (NIH; TR001100). D. Klarin is supported by the National Heart, Lung, and Blood Institute (NHLBI) of NIH under award number T32 HL007734. P. Natarajan is supported by the John S. LaDue Memorial Fellowship in Cardiology from Harvard Medical School. S. Kathiresan is supported by the Ofer and Shelly Nemirovsky Research Scholar Award from the Massachusetts General Hospital, the Donovan Family Foundation, and R01 HL127564. Exome sequencing in ATVB (Atherosclerosis Thrombosis and Vascular Biology), DHM (Deutsches Herzzentrum München Myocardial Infarction Study), JHS (Jackson Heart Study), OHS (Ottawa Heart Study), PROCARDIS (Precocious Coronary Artery Disease Study), and PROMIS (Pakistan Risk of Myocardial Infarction Study) was supported by 5U54HG003067 (to E.S. Lander and S. Gabriel). The JHS is supported and conducted in collaboration with Jackson State University (HHSN268201300049C and HHSN268201300050C), Tougaloo College (HHSN268201300048C), and the University of Mississippi Medical Center (HHSN268201300046C and HHSN268201300047C) contracts from the NHLBI and the National Institute for Minority Health and Health Disparities. Samples for the Leicester study were collected as part of projects funded by the British Heart Foundation (British Heart Foundation Family Heart Study, RG2000010; UK Aneurysm Growth Study, CS/14/2/30841) and the National Institute for Health Research (NIHR Leicester Cardiovascular Biomedical Research Unit Biomedical Research Informatics Centre for Cardiovascular Science, IS_BRU_0211_20033). The DiscovEHR (DiscovEHR project of the Regeneron Genetics Center and the Geisinger Health System) project is funded by Regeneron Pharmaceuticals.

Disclosures

S. Kathiresan has received grants from Bayer Healthcare, Aegerion Pharmaceuticals, and Regeneron Pharmaceuticals; and consulting fees from Merck, Novartis, Sanofi, AstraZeneca, Alnylam Pharmaceuticals, Leerink Partners, Noble Insights, Quest Diagnostics, Genomics PLC, and Eli Lilly and Company; and holds equity in San Therapeutics and Catabasis Pharmaceuticals. The other authors report no conflicts.

Appendix

From the Center for Genomic Medicine, Cardiovascular Research Center, Massachusetts General Hospital, Harvard Medical School, Boston (A.N., H.-H.W., A.V.K., C.A.E., D.K., P.N., S.K.): Program in Medical and Population Genetics, Broad Institute, Cambridge, MA (A.N., H.-H.W., A.V.K., C.A.E., D.K., P.N., S.M.Z., N.G., E.S.L., E.S.L., S.G., S.K.); Cardiovascular and Internal Medicine, Kanazawa University Graduate School of Medicine, Japan (A.N., H.T., M.a.K., A.I., M.Y.); Samsung Advanced Institute for Health Sciences and Technology, Sungkyunkwan University, Samsung Medical Center, Seoul, Republic of Korea (H.-H.W.); Gene Diagnostics and Therapeutics, Research Institute, National Center for Global Health and Medicine, Tokyo, Japan (F.T., N.K.); Center for Integrated Medical Sciences, RIKEN, Yokohama, Japan (K.I., Y.M., M.K.); Regeneron Genetics Center, Tarrytown, NY (S.M., I.B., T.M.T., F.E.D.); Department of Biostatistics, Boston University School of Public Health, MA (G.M.P.); Department of Biomedical Sciences, Humanitas University and Humanitas Clinical and Research Center, Milan, Italy (R.A., S.D.); Ospedale Niguarda, Milan, Italy (P.A.M.); Department of Medicine, Jackson Heart Study(A.C.) and Department of Physiology and Biophysics (J.G.W.), University of Mississippi Medical Center, Jackson; Deutsches Herzzentrum München, Technische Universität München, Deutsches Zentrum für Herz-Kreislauf-Forschung, München, Germany (T.K., H.S.); Munich Heart Alliance, München, Germany (T.K.); Departments of Cardiovascular Sciences and NIHR Leicester Cardiovascular Biomedical Research Unit, University of Leicester, United Kingdom (M.J.B., P.S.B., N.J.S.); Leeds Institute of Cardiovascular and Metabolic Medicine, Leeds University, United Kingdom (A.S.H.); Geisinger Health System, Danville, PA (D.J.C., M.F.M., H.L.K., J.B.L., D.R.L., J.N.M., D.N.H.); Cardiovascular

Epidemiology and Genetics, IMIM (Hospital del Mar Research Institute), Barcelona, Spain (J.M., R.E.); University of Ottawa Heart Institute, Canada (R.M.); Cardiovascular Medicine, Radcliffe Department of Medicine and the Wellcome Trust Centre for Human Genetics, University of Oxford, United Kingdom (M.F., H.W.); Cardiovascular Center and Division of Cardiology, Department of Internal Medicine, National Taiwan University Hospital, Taipei City (J.-M.J.J.); National Taiwan University College of Medicine, Taipei (J.-M.J.J.); Institute of Population Health Sciences, National Health Research Institutes, Miaoli, Taiwan (C.A.H.); Division of Endocrinology and Metabolism, Department of Internal Medicine, Taichung Veterans General Hospital, Taiwan (S.-Y.L., J.-S.W.); Taichung Veterans General Hospital, Taiwan (S.-Y.L.); School of Medicine, National Yang-Ming University, Taipei, Taiwan (J.-S.W.); Departments of Clinical Gene Therapy and Geriatric Medicine and Nephrology, Osaka University Graduate School of Medicine, Suita, Japan (T.K.); Division of Endocrinology and Diabetes, Department of Internal Medicine, Nagoya University Graduate School of Medicine, Japan (E.N.); Department of Diabetes and Endocrinology, Chubu Rosai Hospital, Nagoya, Japan (E.N.); Center for Advanced Medicine and Clinical Research, Nagoya University Hospital, Japan (M.N.); Department of Medical Chemistry, Kurume University School of Medicine, Japan (K.Y.); Department of Genome Science, Aichi-Gakuin University School of Dentistry, Nagoya, Japan (M.Y.); Institute for Translational Genomics and Population Sciences, Los Angeles BioMedical Research Institute, Torrance, CA (J.I.R.); Departments of Medicine and Pediatrics, Harbor-UCLA Medical Center, Torrance, CA (J.I.R.); Department of Genetics (D.J.R.) and Department of Biostatistics and Epidemiology, Perelman School of Medicine (D.S.), University of Pennsylvania, Philadelphia; Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, United Kingdom (J.D.); Cambridge and National Institute for Health Research Blood and Transplant Research Unit in Donor Health and Genomics, Department of Public Health and Primary Care, University of Cambridge, United Kingdom (J.D.); Department of Cardiology, Azienda Ospedaliero-Universitaria di Parma, University of Parma, Italy (D.A.); ASTC: Associazione per lo Studio Della Trombosi in Cardiologia, Pavia, Italy (D.A.); Departments of Computational Medicine and Bioinformatics, Human Genetics, and Internal Medicine, University of Michigan, Ann Arbor (C.J.W.); and Department of Biostatistics, Center for Statistical Genetics, University of Michigan School of Public Health, Ann Arbor (G.R.A.).

Nomura et al

References

- Barter PJ, Caulfield M, Eriksson M, et al; ILLUMINATE Investigators. Effects of torcetrapib in patients at high risk for coronary events. N Engl J Med. 2007;357:2109–2122. doi: 10.1056/NEJMoa0706628.
- Schwartz GG, Olsson AG, Abt M, et al; dal-OUTCOMES Investigators. Effects of dalcetrapib in patients with a recent acute coronary syndrome. N Engl J Med. 2012;367:2089–2099. doi: 10.1056/NEJMoa1206797.
- Nicholls SJ. Evacetrapib fails to reduce major adverse cardiovascular events. ACC16. https://www.acc.org/about-acc/press-releases/2016/04/03/13/02/evacetrapib-fails-to-reduce-major-adverse-cardiovascular-events. Accessed December 16, 2016.
- Schaefer EJ. Effects of cholesteryl ester transfer protein inhibitors on human lipoprotein metabolism: why have they failed in lowering coronary heart disease risk? Curr Opin Lipidol. 2013;24:259–264. doi: 10.1097/MOL.0b013e3283612454.
- Yamashita S, Matsuzawa Y. Re-evaluation of cholesteryl ester transfer protein function in atherosclerosis based upon genetics and pharmacological manipulation. *Curr Opin Lipidol*. 2016;27:459–472. doi: 10.1097/ MOL.0000000000000332.
- Kathiresan S. Will cholesteryl ester transfer protein inhibition succeed primarily by lowering low-density lipoprotein cholesterol? Insights from human genetics and clinical trials. *J Am Coll Cardiol*. 2012;60:2049–2052. doi: 10.1016/j.jacc.2012.08.967.
- Cannon CP, Shah S, Dansky HM, Davidson M, Brinton EA, Gotto AM, Stepanavage M, Liu SX, Gibbons P, Ashraf TB, Zafarino J, Mitchel Y, Barter P; Determining the Efficacy and Tolerability Investigators. Safety of anacetrapib in patients with or at high risk for coronary heart disease. N Engl J Med. 2010;363:2406–2415. doi: 10.1056/NEJMoa1009744.
- Cohen JC, Boerwinkle E, Mosley TH Jr, Hobbs HH. Sequence variations in PCSK9, low LDL, and protection against coronary heart disease. N Engl J Med. 2006;354:1264–1272. doi: 10.1056/NEJMoa054013.

- The Myocardial Infarction Genetics Consortium Investigators, Stitziel NO, Won HH, et al. Inactivating mutations in NPC1L1 and protection from coronary heart disease. N Engl J Med. 2014;371:2072–2082.
- Kathiresan S; Myocardial Infarction Genetics Consortium. A PCSK9 missense variant associated with a reduced risk of early-onset myocardial infarction. N Engl J Med. 2008;358:2299–2300. doi: 10.1056/ NEJMc0707445.
- Moriyama Y, Okamura T, Inazu A, Doi M, Iso H, Mouri Y, Ishikawa Y, Suzuki H, Iida M, Koizumi J, Mabuchi H, Komachi Y. A low prevalence of coronary heart disease among subjects with increased high-density lipoprotein cholesterol levels, including those with plasma cholesteryl ester transfer protein deficiency. *Prev Med.* 1998;27:659–667. doi: 10.1006/ pmed.1998.0340.
- Curb JD, Abbott RD, Rodriguez BL, Masaki K, Chen R, Sharp DS, Tall AR. A prospective study of HDL-C and cholesteryl ester transfer protein gene mutations and the risk of coronary heart disease in the elderly. *J Lipid Res*. 2004;45:948–953. doi: 10.1194/jlr.M300520-JLR200.
- Thompson A, Di Angelantonio E, Sarwar N, Erqou S, Saleheen D, Dullaart RP, Keavney B, Ye Z, Danesh J. Association of cholesteryl ester transfer protein genotypes with CETP mass and activity, lipid levels, and coronary risk. *JAMA*. 2008;299:2777–2788. doi: 10.1001/jama.299.23.2777.
- 14. Hirano K, Yamashita S, Nakajima N, Arai T, Maruyama T, Yoshida Y, Ishigami M, Sakai N, Kameda-Takemura K, Matsuzawa Y. Genetic cholesteryl ester transfer protein deficiency is extremely frequent in the Omagari area of Japan. Marked hyperalphalipoproteinemia caused by CETP gene mutation is not associated with longevity. Arterioscler Thromb Vasc Biol. 1997;17:1053–1059.
- Inazu A, Brown ML, Hesler CB, Agellon LB, Koizumi J, Takata K, Maruhama Y, Mabuchi H, Tall AR. Increased high-density lipoprotein levels caused by a common cholesteryl-ester transfer protein gene mutation. N Engl J Med. 1990;323:1234–1238. doi: 10.1056/NEJM199011013231803.
- Webb TR, Erdmann J, Stirrups KE, et al; Wellcome Trust Case Control Consortium; MORGAM Investigators; Myocardial Infarction Genetics and CARDIOGRAM Exome Consortia Investigators. Systematic evaluation of pleiotropy identifies 6 further loci associated with coronary artery disease. J Am Coll Cardiol. 2017;69:823–836. doi: 10.1016/j. jacc.2016.11.056.
- The TG and HDL Working Group of the Exome Sequencing Project, National Heart, Lung, and Blood Institute. Loss-of-function mutations in APOC3, triglycerides, and coronary disease. N Engl J Med. 2014;371:22–31.
- Cannon CP, Blazing MA, Giugliano RP, et al; IMPROVE-IT Investigators. Ezetimibe added to statin therapy after acute coronary syndromes. N Engl J Med. 2015;372:2387–2397. doi: 10.1056/NEJMoa1410489.
- Sabatine MS, Giugliano RP, Keech AC, Honarpour N, Wiviott SD, Murphy SA, Kuder JF, Wang H, Liu T, Wasserman SM, Sever PS, Pedersen TR; FOURIER Steering Committee and Investigators. Evolocumab and clinical outcomes in patients with cardiovascular disease. N Engl J Med. 2017;376:1713–1722. doi: 10.1056/NEJMoa1615664.
- Dewey FE, Gusarova V, O'Dushlaine C, et al. Inactivating variants in ANGPTL4 and risk of coronary artery disease. N Engl J Med. 2016;374:1123–1133. doi: 10.1056/NEJMoa1510926.
- Assimes TL, Lee IT, Juang JM, et al. Genetics of coronary artery disease in Taiwan: a Cardiometabochip Study by the TAICHI consortium. *PLoS One*. 2016;11:e0138014. doi: 10.1371/journal.pone.0138014.
- Atherosclerosis, Thrombosis, and Vascular Biology Italian Study Group. No evidence of association between prothrombotic gene polymorphisms and the development of acute myocardial infarction at a young age. Circulation. 2003;107:1117–1122.
- Do R, Stitziel NO, Won HH, et al; NHLBI Exome Sequencing Project. Exome sequencing identifies rare LDLR and APOA5 alleles conferring risk for myocardial infarction. *Nature*. 2015;518:102–106. doi: 10.1038/ nature13917.
- The atherosclerosis risk in communities (ARIC) study: design and objectives. The ARIC Investigators. Am J Epidemiol. 1989;129:687–702.
- Taylor HA, Jr. The Jackson Heart Study: an overview. Ethn Dis. 2005;15:S6-1–3.
- Samani NJ, Erdmann J, Hall AS, et al; WTCCC and the Cardiogenics Consortium. Genomewide association analysis of coronary artery disease. N Engl J Med. 2007;357:443–453. doi: 10.1056/NEJMoa072366.
- Myocardial Infarction Genetics and CARDIoGRAM Exome Consortia Investigators. Coding variation in ANGPTL4, LPL, and SVEP1 and the risk of coronary disease. N Engl J Med. 2016;374:1134–1144.
- McPherson R, Pertsemlidis A, Kavaslar N, Stewart A, Roberts R, Cox DR, Hinds DA, Pennacchio LA, Tybjaerg-Hansen A, Folsom AR, Boerwinkle E, Hobbs HH, Cohen JC. A common allele on chromosome 9 associated

- with coronary heart disease. Science. 2007;316:1488–1491. doi: 10.1126/science.1142447.
- Clarke R, Peden JF, Hopewell JC, et al; PROCARDIS Consortium. Genetic variants associated with Lp(a) lipoprotein level and coronary disease. N Engl J Med. 2009;361:2518–2528. doi: 10.1056/NEJMoa0902604.
- Saleheen D, Zaidi M, Rasheed A, et al. The Pakistan Risk of Myocardial Infarction Study: a resource for the study of genetic, lifestyle and other determinants of myocardial infarction in South Asia. Eur J Epidemiol. 2009;24:329–338. doi: 10.1007/s10654-009-9334-y.
- Sentí M, Tomás M, Marrugat J, Elosua R; REGICOR Investigators. Paraoxonase1-192 polymorphism modulates the nonfatal myocardial infarction risk associated with decreased HDLs. Arterioscler Thromb Vasc Biol. 2001;21:415–420.
- Inazu A, Jiang XC, Haraki T, Yagi K, Kamon N, Koizumi J, Mabuchi H, Takeda R, Takata K, Moriyama Y. Genetic cholesteryl ester transfer protein deficiency caused by two prevalent mutations as a major determinant of increased levels of high density lipoprotein cholesterol. *J Clin Invest*. 1994;94:1872–1882. doi: 10.1172/JCI117537.
- 33. Konta A, Ozaki K, Sakata Y, Takahashi A, Morizono T, Suna S, Onouchi Y, Tsunoda T, Kubo M, Komuro I, Eishi Y, Tanaka T. A functional SNP in FLT1 increases risk of coronary artery disease in a Japanese population. *J Hum Genet*. 2016;61:435–441. doi: 10.1038/jhg.2015.171.
- 34. Takeuchi F, Isono M, Katsuya T, Yokota M, Yamamoto K, Nabika T, Shimokawa K, Nakashima E, Sugiyama T, Rakugi H, Yamaguchi S, Ogihara T, Yamori Y, Kato N. Association of genetic variants influencing lipid levels with coronary artery disease in Japanese individuals. *PLoS One*. 2012;7:e46385. doi: 10.1371/journal.pone.0046385.
- Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem. 1972;18:499–502.
- Warnick GR, Knopp RH, Fitzpatrick V, Branson L. Estimating low-density lipoprotein cholesterol by the Friedewald equation is adequate for classifying patients on the basis of nationally recommended cutpoints. *Clin Chem.* 1990;36:15–19.
- Khera AV, Won HH, Peloso GM, et al. Diagnostic yield and clinical utility of sequencing familial hypercholesterolemia genes in patients with severe hypercholesterolemia. *J Am Coll Cardiol*. 2016;67:2578–2589. doi: 10.1016/j.jacc.2016.03.520.
- DePristo MA, Banks E, Poplin R, et al. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat Genet*. 2011;43:491–498. doi: 10.1038/ng.806.
- McLaren W, Pritchard B, Rios D, Chen Y, Flicek P, Cunningham F. Deriving the consequences of genomic variants with the Ensembl API and

- SNP effect predictor. *Bioinformatics*. 2010;26:2069–2070. doi: 10.1093/bioinformatics/btq330.
- Momozawa Y, Akiyama M, Kamatani Y, et al. Low-frequency coding variants in CETP and CFB are associated with susceptibility of exudative agerelated macular degeneration in the Japanese population. *Hum Mol Genet*. 2016;25:5027–5034. doi: 10.1093/hmg/ddw335.
- Global Lipids Genetics Consortium, Willer CJ, Schmidt EM, et al. Discovery and refinement of loci associated with lipid levels. *Nat Genet*. 2013;45:1274–1283.
- Ridker PM, Paré G, Parker AN, Zee RY, Miletich JP, Chasman DI. Polymorphism in the CETP gene region, HDL cholesterol, and risk of future myocardial infarction: genomewide analysis among 18 245 initially healthy women from the Women's Genome Health Study. Circ Cardiovasc Genet. 2009;2:26–33. doi: 10.1161/CIRCGENETICS. 108.817304.
- Voight BF, Peloso GM, Orho-Melander M, et al. Plasma HDL cholesterol and risk of myocardial infarction: a Mendelian randomisation study. *Lancet*. 2012;380:572–580. doi: 10.1016/S0140-6736(12)60312-2.
- Johannsen TH, Frikke-Schmidt R, Schou J, Nordestgaard BG, Tybjærg-Hansen A. Genetic inhibition of CETP, ischemic vascular disease and mortality, and possible adverse effects. *J Am Coll Cardiol*. 2012;60:2041– 2048. doi: 10.1016/j.jacc.2012.07.045.
- Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. Science. 1986;232:34–47.
- Tanigawa H, Billheimer JT, Tohyama J, Zhang Y, Rothblat G, Rader DJ. Expression of cholesteryl ester transfer protein in mice promotes macrophage reverse cholesterol transport. *Circulation*. 2007;116:1267–1273. doi: 10.1161/CIRCULATIONAHA.107.704254.
- 47. Khera AV, Wolfe ML, Cannon CP, Qin J, Rader DJ. On-statin cholesteryl ester transfer protein mass and risk of recurrent coronary events (from the pravastatin or atorvastatin evaluation and infection therapy-thrombolysis in myocardial infarction 22 [PROVE IT-TIMI 22] study). Am J Cardiol. 2010;106:451–456. doi: 10.1016/j.amjcard.2010.03.057.
- Tall AR, Yvan-Charvet L, Terasaka N, Pagler T, Wang N. HDL, ABC transporters, and cholesterol efflux: implications for the treatment of atherosclerosis. *Cell Metab*. 2008;7:365–375. doi: 10.1016/j. cmet.2008.03.001.
- Brousseau ME, Diffenderfer MR, Millar JS, Nartsupha C, Asztalos BF, Welty FK, Wolfe ML, Rudling M, Björkhem I, Angelin B, Mancuso JP, Digenio AG, Rader DJ, Schaefer EJ. Effects of cholesteryl ester transfer protein inhibition on high-density lipoprotein subspecies, apolipoprotein A-I metabolism, and fecal sterol excretion. *Arterioscler Thromb Vasc Biol.* 2005;25:1057–1064. doi: 10.1161/01.ATV.0000161928.16334.dd.